concd to 200 ml under red. pres. and extracted with hot distilled H_2O . The H_2O soluble portion was treated successively with CHCl₃ and EtOAc, concd and chromatographed on silica gel. Elution with MeOH yielded an amorphous compound which crystallized from MeOH and when further purified by HPLC afforded crystals, mp 188–189° (Found: C, 48.42; H, 5.15. $C_{11}H_{14}O_8$ requires C, 48.17; H, 5.11%); $[\alpha]_D^{20}$ +162° (H_2O); slightly bitter in taste. Compound 1 is soluble in H_2O , sparingly soluble in cold EtOH, and insoluble in Et_2O . It gave no colour with NaOH or FeCl₃ soln, did not absorb Br_2 – H_2O but gave a positive Molisch test. IR ν_{max}^{KBr} cm⁻¹; 3550–3320, 3100, 1662, 1610, 1240, 1150–1130, 780, 770 and 840; ¹H NMR (90 MHz) (D_2O): δ 3.7–3.8 (m, 4H, H-2', H-3', H-4', H-5'), 3.98 and 4.05 (2H, H-6'), 5.1 (1H, H-1'), 6.8 (d, 1H, H-5), 8.32 (d, 1H, H-6) and 8.5 (s, 1H, H-2); MS m/z: 275 [M+1] +, 183, 163, 162, 145, 141, 86, 85, 84, 73, 60, 57 and 55.

Glucoside tetra-acetate. Compound 1 on acetylation (Ac₂O-pyridine) and recrystallization from MeOH gave a glucoside tetra-acetate; MS m/z (rel. int.): 442 [M]⁺, 169 (100), 331 (87), 109 (62), 211 (30), 271 (22), 42 (13) and 112 (11).

Acid hydrolysis of 1. Acid hydrolysis (6% methanolic H_2SO_4 , 5 hr) gave only a small quantity of white amorphous aglycone from the EtOAc extract, which gave a red colour with FeCl₃. MS m/z: 112 [M]⁺; IR ν_{max}^{KBr} cm⁻¹: 1662 and 1610 (diagnostic bands

of γ-pyrone). The remaining aq. layer reduced Fehlings soln and Tollen's reagent, and the sugar was identified as p-glucose by co-PC in n-BuOH-HOAc-H₂O (4:1:5), EtOAc-pyridine-H₂O (5:2:7), n-BuOH-pyridine-H₂O (6:4:3) and EtOAc-HOAc-H₂O (5:2:2).

Acknowledgements—We are thankful to Dr. D. S. Bhakuni and the authorities of C.D.R.I., Lucknow for spectral data and to Mr. P. S. Green, The Royal Botanic Gardens, Kew for the verification of the plant material.

REFERENCES

- 1. Plouvier, V. (1964) Compt. Rend. 258, 1099.
- Renwei, Z., Shengyuan, Y and Yongyue, L. (1981) Acta Pharm. Sinica 16, 68
- Nakanishi, K. and Solomon, P. H. (1977) Infrared Absorption Spectroscopy, 2nd edn, p. 249. Holden-Day, San Francisco.
- McGillivray, D. L. and Poulton, G. A. (1978) Org. Mass Spectrom. 13, 296.
- Biemann, K., DeJongh, D. C. and Schnoes, H. K. (1963) J. Am Chem. Soc. 85, 1763.
- 6. Pearl, I. A. and Darling, S. F. (1968) Phytochemistry 7, 831.

Phytochemistry, Vol 23, No 9, pp 2091-2093, 1984. Printed in Great Britain.

0031-9422/84 \$3 00 + 0 00 © 1984 Pergamon Press Ltd

3-BENZYL-4-CHROMANONES FROM MUSCARI COMOSUM

MATTEO ADINOLFI, GASPARE BARONE, MARGHERITA BELARDINI*, ROSA LANZETTA, GUGLIELMO LAONIGRO and MICHELANGELO PARRILLI

Istituto di Chimica Organica e Biologica, Università di Napoli, Via Mezzocannone 16, 80134 Napoli, Italy; *Istituto di Chimica Applicata, Università di Napoli, Piazzale Tecchio, Napoli, Italy

(Received 27 January 1984)

Key Word Index—*Muscari comosum*; Liliaceae, 3-benzyl-4-chromanones; homoisoflavanones; 5-hydroxy-3-(*p*-hydroxybenzyl)-7,8-dimethoxy-4-chromanone; 5,8-dihydroxy-3-(*p*-hydroxybenzyl)-7-methoxy-4-chromanone; 5,7-dihydroxy-3-(*p*-hydroxybenzyl)-6-methoxy-4-chromanone.

Abstract—From the bulbs of *Muscari comosum* two novel 3-benzyl-4-chromanones, 7-O-methyl-3,9-dihydropunctatin and 8-O-demethyl-7-O-methyl-3,9-didropunctatin, were isolated.

INTRODUCTION

The bulbs of *Muscari comosum* have been shown to be a rich source of both triterpene glycosides [1] and free triterpenes [2]. One of these latter compounds, eucosterol, was found for the first time in some *Eucomis* species of the Liliaceae family [3] which were also shown [4] to contain some members of a new class of natural compounds, 3-benzyl(idene)-4-chromanones (or 'homoisoflavanones'). This prompted us to investigate the occurrence of this type of compound in *M. comosum*. This study led us to isolate two novel 3-benzyl-chromanones,

namely 7-O-methyl-3,9-dihydropunctatin 1 and 8-O-demethyl-7-O-methyl-3,9-dihydropunctatin 2, in addition to the already known [5] 3,9-dihydroeucomnalin 3. The structures of 1 and 2 were elucidated by spectral analysis and chemical correlation.

RESULTS AND DISCUSSION

Compound 1 possesses the molecular formula $C_{18}H_{18}O_6$ (high-resolution mass spectrum). In the ¹H NMR spectrum (Table 1) the signals of the –(2)CH₂–(3)CH–(9)CH₂– grouping were clearly seen; they were easily assigned by comparison to the reported chemical shift values for similar groupings in 3-benzyl-4-chromanones [4]. The presence of a hydroxytropylium

[†]Author to whom correspondence should be addressed.

2092 Short Reports

Table 1.	¹ H NMR	(270 MHz) chemical	shifts in	DMSO-d ₆ *
----------	--------------------	----------	------------	-----------	-----------------------

Compound	C-2	C-3	C-5	C-6	C-7	C-8	C-9	C-2', C-6'	C-3', C-5'	C-4′
1	4.16 m 4.34 m AB of ABX	30m	12 06† s	6.22 s	3.84 <i>s</i>	3.61 s	2.63 m 3.0 m	7.04 $J = 8.54$ AA'BB'	6.70	9.26† s
2	4.12 m 4.20 m AB of ABX	3.0 m	11.83† s	6.19 s	3.84 <i>s</i>	8.16† s	2.62 m 3 0 m		6.72	9.26† s
3	4 06 m 4 23 m AB of ABX	3.0 m	12.24† s	3.69 s	10.48 s	5.97 s	2 60 m 3.0 m		6.72	9.26† s

^{*}All chemical shifts are given in δ (ppm) relative to TMS. Coupling constants are given in Hz.

Table 2. ¹³C NMR (67.88 MHz) chemical shifts of 1, 2 and 3 in DMSO-d₆*

Carbon	1	2	3
2	69.08	68.81	68 83
3	45.70	45.94	45.60
4	198 51	198.54	198.38
4a	101 67	101 64	101 24
5	159 07†	155 87†	155.31†
6	92 78	92.49	128.99‡
7	160 85†	156.83†	159.40†
8	128.68‡	126 28‡	94 66
8a	153 34†	148.02†	157.85†
9	31.12	31 08	31 07
1'	125.93‡	127 98‡	127 94‡
2', 6'	129 92	129.80	129.84
3', 5'	115 15	115 16	115 18
4'	155 59†	155.81†	155.79†
OMe	60.44	55 99	59 89
OMe	56.19		

^{*}Chemical shifts are given in δ (ppm) relative to TMS. The assignments are based on on- and off-resonance spectra and on comparison to data from ref. [4].

Table 3. Nuclear Overhauser effects measured on 1 in DMSO- d_6 *

Irradiation	Observed	
δ6.22 (H-6)	a δ 3.84 (7-OMe)	
` ,	b 12.06 (5-OH)	
12.06 (5-OH)	c 6.22 (H-6)	
3.84 (7-OMe)	d 6 22 (H-6)	
` ,	e 3.61 (8-OMe)	

^{*}The NOE difference FIDs were obtained by gated decoupling

fragment (m/z 107) in the mass spectrum and the ¹H NMR signals of an aromatic AA'BB' system (δ 6.70 and 7.04, J = 8.5 Hz; protons at C-2', C-3', C-5' and C-6') indicate the B-ring substitution pattern. The lowfield signal due to a

hydroxyl proton (δ 12.06) is assigned to the strongly hydrogen-bonded 5-hydroxyl group [5]. Two methoxyl groups are attached to the A ring (δ 3.85 and 3.61), which also carries the proton responsible for the singlet at δ 6.23. According to the above data, the UV spectrum of 1 (EtOH) exhibited a main absorption at 288 nm ($\log \epsilon$ 4.2), which undergoes a bathochromic shift of 30 nm upon addition of aluminum chloride, as expected considering the presence of the 5-hydroxyl [6]; addition of sodium acetate does not cause a similar shift, according to the absence of a hydroxyl group at the 7-position [6].

The attachment sites of the two methoxyl groups at the A ring were identified by examining the ¹³C NMR

[†]Protons exchange with D2O

^{†, ‡}Interchangeable values.

Short Reports 2093

spectrum of 1 (Table 2) and observing that in the fully-coupled spectrum the methine carbon of the A ring (δ 92.78) appears as a doublet ($J_{C,H}^1 = 163 \text{ Hz}$) further split by a $J_{C,H}^2 = 6.6 \text{ Hz}$. This coupling through three bonds can only occur between the 6-carbon and the 5-hydroxyl proton. Accordingly, upon addition of D_2O , in the fully-coupled spectrum the signal appears as a simple doublet [7]. Thus the 6-position of the A ring does not carry a substituent. As a consequence, the methoxyl groups are attached at positions 7 and 8. Further support for structure 1 was achieved from NOE experiments (Table 3). Results a, b and c accord only to the presence of a proton at the 6-position.

Compound 2 possesses the molecular formula C₁₇H₁₆O₆ (high-resolution mass spectrum). ¹H and ¹³CNMR data are summarized in Tables 1 and 2, respectively. The ¹H NMR spectrum displayed the signals of the -(2)CH₂-(3)CH-(9)CH₂- grouping and of the AA'BB' system of the B-ring protons. The B-ring substitution pattern was also deduced from the appearance of the hydroxytropylium peak (m/z 107) in the mass spectrum. The $\delta 11.83$ singlet in the ¹H NMR spectrum must be due to the 5-hydroxyl proton, involved in a strong hydrogen bond. UV absorption (293 nm; EtOH) undergoes a bathochromic shift of 30 nm upon addition of aluminum chloride (presence of the 5-hydroxyl). In the fully-coupled 13C-spectrum of 2 the A-ring methine carbon (δ 92.49) appears to be coupled to the 5-hydroxyl proton ($J_{C,H}^3 = 6.5 \text{ Hz}$). The OMe group must be at C-7 and the third hydroxyl group at C-8 since the UV absorption maximum is not shifted upon addition of sodium acetate (absence of the 7-hydroxyl).

Both 1 and 2 were permethylated by treatment with dimethyl sulphate-potassium carbonate in acetone and yielded the same fully methylated derivative. As expected, mp and ¹H NMR spectra in CDCl₃ and C₆D₆ of these were identical to those described for compound 4, obtained by permethylation of 3,9-dihydropunctatin [8].

Compound 3, mp $207-209^{\circ}$, $C_{17}H_{16}O_{6}$ (highresolution mass spectrum), exhibited an ¹H NMR spectrum (Table 1) which closely resembled those reported for 3,9-dihydroeucomnalin (\equiv 3,9-dihydroautumnalin, 3) [5] and for 3,9-dihydropunctatin 5 [8]. However, compound 3 and 3,9-dihydroeucomnalin were shown to be identical by the fact that the methine carbon of the A ring appears in the fully-coupled 13C spectrum as a simple doublet. This was confirmed by conversion (dimethyl sulphate-potassium carbonate in acetone) into the fully methylated derivative 6, whose mp and ¹H NMR spectra in CDCl₃ and in C₆D₆ were identical to those described for permethylated 3,9-dihydroeucomnalin [5] and different from those described for permethylated 3,9-dihydropunctatin [8]. The as yet unreported ¹³C-spectrum of 3 is summarized in Table 2.

EXPERIMENTAL

Isolation of 3-benzylchroman-4-ones. Fresh bulbs (1 kg) of M. comosum Mill. (Liliaceae) (collected in the autumn in Puglia,

Italy, and authenticated by the Botanical Garden of the University of Naples) were homogenized in a mechanical stirrer, freeze-dried and extracted in a Soxhlet apparatus with petrol (12 hr) and then with Et₂O (12 hr). The Et₂O extract was evapd (3 g) and chromatographed on a silica gel (90 g) column with hexane containing increasing proportions of Et₂O. The fraction (2 g) eluted with Et₂O was chromatographed on a silica gel (60 g) column with CHCl₃-EtOAc. The fraction (0.5 g) eluted with CHCl₃-EtOAc (19.1) was chromatographed on a silica gel (15 g) column with C₆H₆-EtOAc. Five fractions were collected: a (50 mg), b (75 mg), c (150 mg), d (85 mg), and e (200 mg) (increasing polarity order).

Prep. TLC (silica gel, C_6H_6 -Et₂O (7.3), 2 runs) of fraction *a* yielded compound 1 (30 mg) as a vitreous solid. EIMS, 70 eV, m/z (rel. int.). 330.1113 ([M]⁺; calc. for $C_{18}H_{18}O_6$ 330.1103) (40), 107 (100).

Crystallization of fraction e from CHCl₃ gave compound 2 (60 mg), mp 172–174°. EIMS, 70 eV, m/z (rel. int.): 316.0959 ([M]⁺; calc for. $C_{17}H_{16}O_6$ 316.0947) (45); 107 (100).

Crystallization of fraction c from CHCl₃ gave compound 3 (80 mg), mp 209°. EIMS, 70 eV, m/z (rel. int.) 316.0962 ([M]⁺; calc. for $C_{17}H_{16}O_6$ 316.0947) (45); 107 (100).

Methyl derivatives 4 and 6. Separate methylation of samples of 1 and 2 with Me_2SO_4 – K_2CO_3 in dry Me_2CO (room temp, 24 hr) [9] gave Me derivative 4, mp 99–100°, in both cases The product was identical (1H NMR in CDCl₃ and in C_6D_6 ; mp) to the methylation product of 3,9-dihydropunctatin [5].

Methylation of 3 using the above conditions gave 6, mp 75–76°, identical (mp; ${}^{1}H$ NMR in CDCl₃ and in C₆D₆) to the Me derivative of 3,9-dihydroeucomnalin [8].

Acknowledgements—This work was supported by Ministero della Pubblica Istruzione. NMR spectra were performed at the Centro di Metodologie Chimico-Fisiche della Università di Napoli.

REFERENCES

- Adınolfi, M., Barone, G., Lanzetta, R., Laonigro, G., Mangoni, L. and Parrilli, M. (1984) Can J. Chem. 62 (in press) and references cited therein.
- Adinolfi, M., Barone, G., Lanzetta, R., Laonigro, G., Mangoni, L and Parrilli, M. (1984) J. Nat. Prod. 47 (in press) and references cited therein
- 3 Ziegler, R. and Tamm, C. (1976) Helv. Chim. Acta 59, 1997.
- Heller, W. and Tamm, C. (1981) Fortschr. Chem. Org. Naturst. 40, 105.
- 5 Sidwell, W. T. L. and Tamm, C. (1970) Tetrahedron Letters 475.
- Markham, K. R., and Mabry, T. J. (1975) in The Flavonoids (Harborne, J. B., Mabry, T. J. and Mabry, H., eds) p. 45. Chapman & Hall, London.
- 7 Wehrli, F. W. (1975) Chem. Commun. 663.
- 8. Finckh, R. E and Tamm, C. (1970) Experientia 26, 472.
- Heller, W., Andermatt, P., Schaad, W A. and Tamm, C (1976) Helv Chim. Acta 59, 2048.