concd to 200 ml under red. pres. and extracted with hot distilled  $H_2O$ . The  $H_2O$  soluble portion was treated successively with CHCl<sub>3</sub> and EtOAc, concd and chromatographed on silica gel. Elution with MeOH yielded an amorphous compound which crystallized from MeOH and when further purified by HPLC afforded crystals, mp 188–189° (Found: C, 48.42; H, 5.15.  $C_{11}H_{14}O_8$  requires C, 48.17; H, 5.11%);  $[\alpha]_D^{20}$  +162° ( $H_2O$ ); slightly bitter in taste. Compound 1 is soluble in  $H_2O$ , sparingly soluble in cold EtOH, and insoluble in  $Et_2O$ . It gave no colour with NaOH or FeCl<sub>3</sub> soln, did not absorb  $Br_2$ – $H_2O$  but gave a positive Molisch test. IR  $\nu_{max}^{KBr}$  cm<sup>-1</sup>; 3550–3320, 3100, 1662, 1610, 1240, 1150–1130, 780, 770 and 840; <sup>1</sup>H NMR (90 MHz) ( $D_2O$ ):  $\delta$ 3.7–3.8 (m, 4H, H-2', H-3', H-4', H-5'), 3.98 and 4.05 (2H, H-6'), 5.1 (1H, H-1'), 6.8 (d, 1H, H-5), 8.32 (d, 1H, H-6) and 8.5 (s, 1H, H-2); MS m/z: 275 [M+1] +, 183, 163, 162, 145, 141, 86, 85, 84, 73, 60, 57 and 55.

Glucoside tetra-acetate. Compound 1 on acetylation (Ac<sub>2</sub>O-pyridine) and recrystallization from MeOH gave a glucoside tetra-acetate; MS m/z (rel. int.): 442 [M]<sup>+</sup>, 169 (100), 331 (87), 109 (62), 211 (30), 271 (22), 42 (13) and 112 (11).

Acid hydrolysis of 1. Acid hydrolysis (6% methanolic  $H_2SO_4$ , 5 hr) gave only a small quantity of white amorphous aglycone from the EtOAc extract, which gave a red colour with FeCl<sub>3</sub>. MS m/z: 112 [M]<sup>+</sup>; IR  $\nu_{max}^{KBr}$  cm<sup>-1</sup>: 1662 and 1610 (diagnostic bands

of γ-pyrone). The remaining aq. layer reduced Fehlings soln and Tollen's reagent, and the sugar was identified as p-glucose by co-PC in n-BuOH-HOAc-H<sub>2</sub>O (4:1:5), EtOAc-pyridine-H<sub>2</sub>O (5:2:7), n-BuOH-pyridine-H<sub>2</sub>O (6:4:3) and EtOAc-HOAc-H<sub>2</sub>O (5:2:2).

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# 3-BENZYL-4-CHROMANONES FROM MUSCARI COMOSUM

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**Key Word Index**—*Muscari comosum*; Liliaceae, 3-benzyl-4-chromanones; homoisoflavanones; 5-hydroxy-3-(*p*-hydroxybenzyl)-7,8-dimethoxy-4-chromanone; 5,8-dihydroxy-3-(*p*-hydroxybenzyl)-7-methoxy-4-chromanone; 5,7-dihydroxy-3-(*p*-hydroxybenzyl)-6-methoxy-4-chromanone.

**Abstract**—From the bulbs of *Muscari comosum* two novel 3-benzyl-4-chromanones, 7-O-methyl-3,9-dihydropunctatin and 8-O-demethyl-7-O-methyl-3,9-didropunctatin, were isolated.

### INTRODUCTION

The bulbs of *Muscari comosum* have been shown to be a rich source of both triterpene glycosides [1] and free triterpenes [2]. One of these latter compounds, eucosterol, was found for the first time in some *Eucomis* species of the Liliaceae family [3] which were also shown [4] to contain some members of a new class of natural compounds, 3-benzyl(idene)-4-chromanones (or 'homoisoflavanones'). This prompted us to investigate the occurrence of this type of compound in *M. comosum*. This study led us to isolate two novel 3-benzyl-chromanones,

namely 7-O-methyl-3,9-dihydropunctatin 1 and 8-O-demethyl-7-O-methyl-3,9-dihydropunctatin 2, in addition to the already known [5] 3,9-dihydroeucomnalin 3. The structures of 1 and 2 were elucidated by spectral analysis and chemical correlation.

## RESULTS AND DISCUSSION

Compound 1 possesses the molecular formula  $C_{18}H_{18}O_6$  (high-resolution mass spectrum). In the <sup>1</sup>H NMR spectrum (Table 1) the signals of the –(2)CH<sub>2</sub>–(3)CH–(9)CH<sub>2</sub>– grouping were clearly seen; they were easily assigned by comparison to the reported chemical shift values for similar groupings in 3-benzyl-4-chromanones [4]. The presence of a hydroxytropylium

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Table 1	1.	<sup>1</sup> H NMR	(270)	MHz) chemica	ıl shifts ın	DMSO-d <sub>6</sub> *
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Compound	C-2	C-3	C-5	C-6	C-7	C-8	C-9	C-2', C-6'	C-3', C-5'	C-4′
1	4.16 m 4.34 m AB of ABX	30m	12 06† s	6.22 s	3.84 <i>s</i>	3.61 s	2.63 m 3.0 m	7.04 $J = 8.54$ $AA'BB'$	6.70	9.26† s
2	4.12 m 4.20 m AB of ABX	3.0 m	11.83† s	6.19 s	3.84 <i>s</i>	8.16† s	2.62 m 3 0 m		6.72	9.26† s
3	4 06 m 4 23 m AB of ABX	3.0 m	12.24† s	3.69 s	10.48 s	5.97 s	2 60 m 3.0 m		6.72	9.26† s

<sup>\*</sup>All chemical shifts are given in  $\delta$  (ppm) relative to TMS. Coupling constants are given in Hz.

Table 2. <sup>13</sup>C NMR (67.88 MHz) chemical shifts of 1, 2 and 3 in DMSO-d<sub>6</sub>\*

Carbon	1	2	3
2	69.08	68.81	68 83
3	45.70	45.94	45.60
4	198 51	198.54	198.38
4a	101 67	101 64	101 24
5	159 07†	155 87†	155.31†
6	92 78	92.49	128.99‡
7	160 85†	156.83†	159.40†
8	128.68‡	126 28‡	94 66
8a	153 34†	148.02†	157.85†
9	31.12	31 08	31 07
1'	125.93‡	127 98‡	127 94‡
2', 6'	129 92	129.80	129.84
3', 5'	115 15	115 16	115 18
4'	155 59†	155.81†	155.79†
OMe	60.44	55 99	59 89
OMe	56.19		

<sup>\*</sup>Chemical shifts are given in  $\delta$  (ppm) relative to TMS. The assignments are based on on- and off-resonance spectra and on comparison to data from ref. [4].

Table 3. Nuclear Overhauser effects measured on 1 in DMSO- $d_6$ \*

Irradiation	Observed		
δ6.22 (H-6)	a δ 3.84 (7-OMe)		
` ,	b 12.06 (5-OH)		
12.06 (5-OH)	c 6.22 (H-6)		
3.84 (7-OMe)	d 6 22 (H-6)		
` ,	e 3.61 (8-OMe)		

<sup>\*</sup>The NOE difference FIDs were obtained by gated decoupling

fragment (m/z 107) in the mass spectrum and the <sup>1</sup>H NMR signals of an aromatic AA'BB' system ( $\delta$ 6.70 and 7.04, J = 8.5 Hz; protons at C-2', C-3', C-5' and C-6') indicate the B-ring substitution pattern. The lowfield signal due to a

hydroxyl proton ( $\delta$ 12.06) is assigned to the strongly hydrogen-bonded 5-hydroxyl group [5]. Two methoxyl groups are attached to the A ring ( $\delta$ 3.85 and 3.61), which also carries the proton responsible for the singlet at  $\delta$ 6.23. According to the above data, the UV spectrum of 1 (EtOH) exhibited a main absorption at 288 nm ( $\log \epsilon$ 4.2), which undergoes a bathochromic shift of 30 nm upon addition of aluminum chloride, as expected considering the presence of the 5-hydroxyl [6]; addition of sodium acetate does not cause a similar shift, according to the absence of a hydroxyl group at the 7-position [6].

The attachment sites of the two methoxyl groups at the A ring were identified by examining the <sup>13</sup>C NMR

<sup>†</sup>Protons exchange with D2O

<sup>†, ‡</sup>Interchangeable values.

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spectrum of 1 (Table 2) and observing that in the fully-coupled spectrum the methine carbon of the A ring ( $\delta$ 92.78) appears as a doublet ( $J_{C,H}^1 = 163 \text{ Hz}$ ) further split by a  $J_{C,H}^2 = 6.6 \text{ Hz}$ . This coupling through three bonds can only occur between the 6-carbon and the 5-hydroxyl proton. Accordingly, upon addition of  $D_2O$ , in the fully-coupled spectrum the signal appears as a simple doublet [7]. Thus the 6-position of the A ring does not carry a substituent. As a consequence, the methoxyl groups are attached at positions 7 and 8. Further support for structure 1 was achieved from NOE experiments (Table 3). Results a, b and c accord only to the presence of a proton at the 6-position.

Compound 2 possesses the molecular formula C<sub>17</sub>H<sub>16</sub>O<sub>6</sub> (high-resolution mass spectrum). <sup>1</sup>H and <sup>13</sup>CNMR data are summarized in Tables 1 and 2, respectively. The <sup>1</sup>H NMR spectrum displayed the signals of the -(2)CH<sub>2</sub>-(3)CH-(9)CH<sub>2</sub>- grouping and of the AA'BB' system of the B-ring protons. The B-ring substitution pattern was also deduced from the appearance of the hydroxytropylium peak (m/z 107) in the mass spectrum. The  $\delta 11.83$  singlet in the <sup>1</sup>H NMR spectrum must be due to the 5-hydroxyl proton, involved in a strong hydrogen bond. UV absorption (293 nm; EtOH) undergoes a bathochromic shift of 30 nm upon addition of aluminum chloride (presence of the 5-hydroxyl). In the fully-coupled 13C-spectrum of 2 the A-ring methine carbon ( $\delta$ 92.49) appears to be coupled to the 5-hydroxyl proton ( $J_{C,H}^3 = 6.5 \text{ Hz}$ ). The OMe group must be at C-7 and the third hydroxyl group at C-8 since the UV absorption maximum is not shifted upon addition of sodium acetate (absence of the 7-hydroxyl).

Both 1 and 2 were permethylated by treatment with dimethyl sulphate-potassium carbonate in acetone and yielded the same fully methylated derivative. As expected, mp and <sup>1</sup>H NMR spectra in CDCl<sub>3</sub> and C<sub>6</sub>D<sub>6</sub> of these were identical to those described for compound 4, obtained by permethylation of 3,9-dihydropunctatin [8].

Compound 3, mp  $207-209^{\circ}$ ,  $C_{17}H_{16}O_{6}$  (highresolution mass spectrum), exhibited an <sup>1</sup>H NMR spectrum (Table 1) which closely resembled those reported for 3,9-dihydroeucomnalin ( $\equiv$ 3,9-dihydroautumnalin, 3) [5] and for 3,9-dihydropunctatin 5 [8]. However, compound 3 and 3,9-dihydroeucomnalin were shown to be identical by the fact that the methine carbon of the A ring appears in the fully-coupled 13C spectrum as a simple doublet. This was confirmed by conversion (dimethyl sulphate-potassium carbonate in acetone) into the fully methylated derivative 6, whose mp and <sup>1</sup>H NMR spectra in CDCl<sub>3</sub> and in C<sub>6</sub>D<sub>6</sub> were identical to those described for permethylated 3,9-dihydroeucomnalin [5] and different from those described for permethylated 3,9-dihydropunctatin [8]. The as yet unreported <sup>13</sup>C-spectrum of 3 is summarized in Table 2.

### **EXPERIMENTAL**

Isolation of 3-benzylchroman-4-ones. Fresh bulbs (1 kg) of M. comosum Mill. (Liliaceae) (collected in the autumn in Puglia,

Italy, and authenticated by the Botanical Garden of the University of Naples) were homogenized in a mechanical stirrer, freeze-dried and extracted in a Soxhlet apparatus with petrol (12 hr) and then with Et<sub>2</sub>O (12 hr). The Et<sub>2</sub>O extract was evapd (3 g) and chromatographed on a silica gel (90 g) column with hexane containing increasing proportions of Et<sub>2</sub>O. The fraction (2 g) eluted with Et<sub>2</sub>O was chromatographed on a silica gel (60 g) column with CHCl<sub>3</sub>-EtOAc. The fraction (0.5 g) eluted with CHCl<sub>3</sub>-EtOAc (19.1) was chromatographed on a silica gel (15 g) column with C<sub>6</sub>H<sub>6</sub>-EtOAc. Five fractions were collected: a (50 mg), b (75 mg), c (150 mg), d (85 mg), and e (200 mg) (increasing polarity order).

Prep. TLC (silica gel,  $C_6H_6$ -Et<sub>2</sub>O (7.3), 2 runs) of fraction *a* yielded compound 1 (30 mg) as a vitreous solid. EIMS, 70 eV, m/z (rel. int.). 330.1113 ([M]<sup>+</sup>; calc. for  $C_{18}H_{18}O_6$  330.1103) (40), 107 (100).

Crystallization of fraction e from CHCl<sub>3</sub> gave compound 2 (60 mg), mp 172–174°. EIMS, 70 eV, m/z (rel. int.): 316.0959 ([M]<sup>+</sup>; calc for.  $C_{17}H_{16}O_6$  316.0947) (45); 107 (100).

Crystallization of fraction c from CHCl<sub>3</sub> gave compound 3 (80 mg), mp 209°. EIMS, 70 eV, m/z (rel. int.) 316.0962 ([M]<sup>+</sup>; calc. for  $C_{17}H_{16}O_6$  316.0947) (45); 107 (100).

Methyl derivatives 4 and 6. Separate methylation of samples of 1 and 2 with  $Me_2SO_4$ – $K_2CO_3$  in dry  $Me_2CO$  (room temp, 24 hr) [9] gave Me derivative 4, mp 99–100°, in both cases The product was identical ( $^1H$  NMR in CDCl<sub>3</sub> and in  $C_6D_6$ ; mp) to the methylation product of 3,9-dihydropunctatin [5].

Methylation of 3 using the above conditions gave 6, mp 75–76°, identical (mp;  ${}^{1}H$  NMR in CDCl<sub>3</sub> and in C<sub>6</sub>D<sub>6</sub>) to the Me derivative of 3,9-dihydroeucomnalin [8].

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